



Azole resistance in *Aspergillus* species in Iran: Is there a challenge coming up?

Mohammad T. Hedayati

Invasive Fungi Research Center/ Department of Medical Mycology, Mazandaran University of Medical Sciences, Sari, Iran



Management of
fungal diseases in

Plants, animals
and humans, and for
material preservation.

Major benefits
in terms of

Food security, animal
health, quality of life,
and survival of patients
with life-threatening
fungal diseases

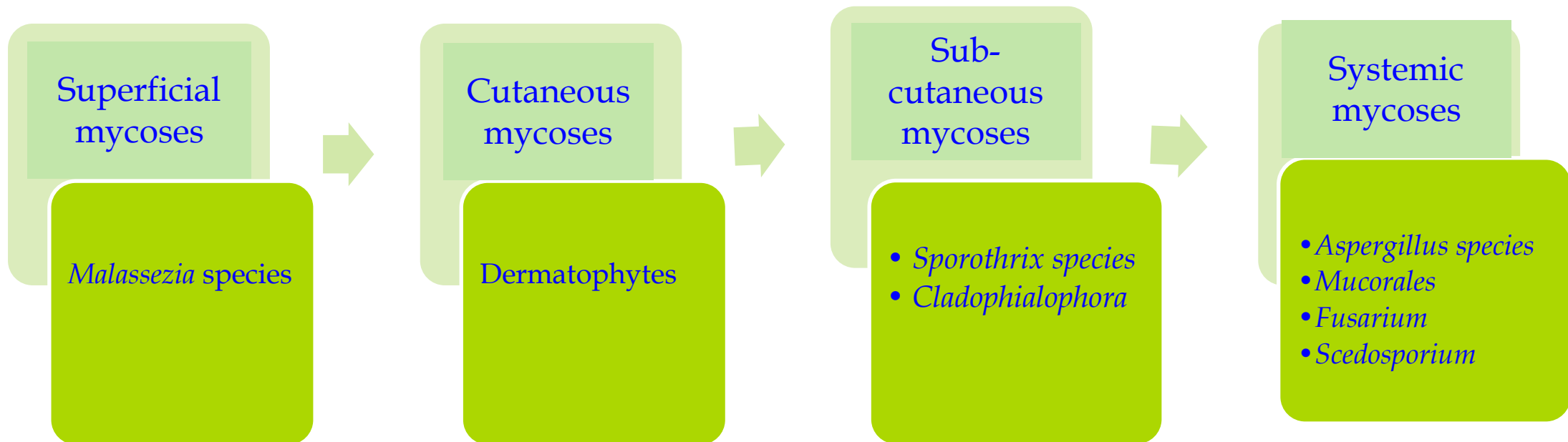
Broad use of the
same antifungal class
comes with a risk

Selection of antifungal
resistance

In vitro assays of
azole susceptibility

Reproducible results that are
predictive of therapeutic
response in experimental
models and patients

Azoles in human:



Critical Priority Group



Cryptococcus neoformans



Aspergillus fumigatus



Candida auris



Candida albicans

WHO fungal priority pathogens list

High Priority Group



Nakaseomyces glabrata
(*Candida glabrata*)



Eumycetoma
causative agents



Fusarium spp.



Candida parapsilosis



Histoplasma spp.



Mucorales



Candida tropicalis

Medium Priority Group



Scedosporium spp.



Lomentospora prolificans



Coccidioides spp.



Pichia kudriavzevii
(*Candida krusei*)



Cryptococcus gattii



Talaromyces marneffei



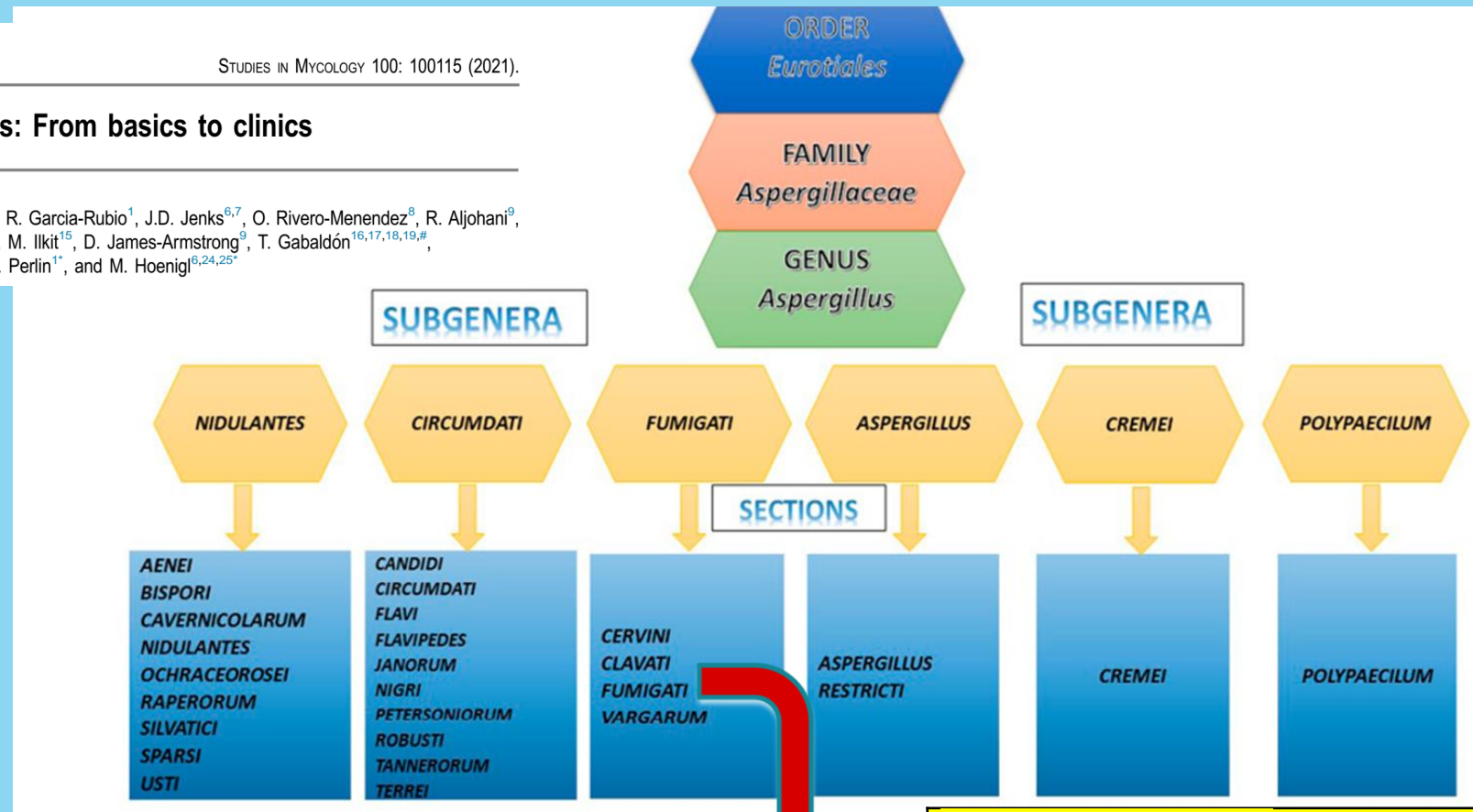
Pneumocystis jirovecii



Paracoccidioides spp.

Aspergillus fumigatus and aspergillosis: From basics to clinics

A. Arastehfar^{1,#}, A. Carvalho^{2,3,#}, J. Houbraken^{4,#}, L. Lombardi⁵, R. Garcia-Rubio¹, J.D. Jenks^{6,7}, O. Rivero-Menendez⁸, R. Aljohani⁹, I.D. Jacobsen^{10,11}, J. Berman¹², N. Oshero¹³, M.T. Hedayati¹⁴, M. Ilkit¹⁵, D. James-Armstrong⁹, T. Gabaldón^{16,17,18,19,#}, J. Meletiadi²⁰, M. Kostrzewa²¹, W. Pan²², C. Lass-Flöri²³, D.S. Perlin^{1*}, and M. Hoernig^{6,24,25*}



Species

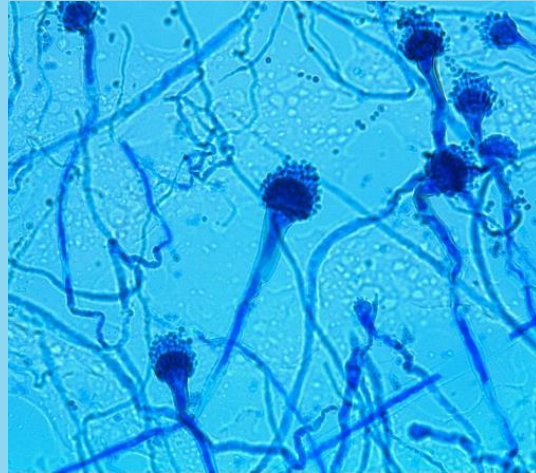
- Aspergillus fumigatus*
- A. udagawae* (*Neosartorya udagawae*)
- A. Thermomutatus* (*Neosartorya pseudofischeri*)
- A. lentulus*
- A. spinosus*
- A. hiratsukae* (*Neosartorya hiratsukae*)
- A. Fischerianus* (*Neosartorya fischeri*)
- A. fumisynnematus*

- A. fumisynnematus*
- A. fumigatiaffinis*
- A. novofumigatus*
- A. laciniosa* (*Neosartorya laciniosa*)
- A. wyomingensis*
- A. pseudoviridinutans*
- A. felis*
- A. parafelis*
- A. pseudofelis*

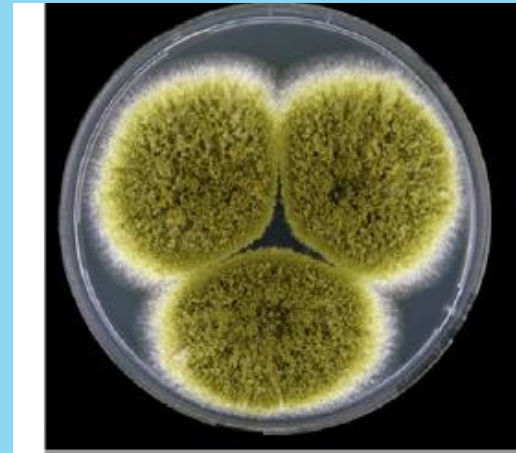
Interactive quiz



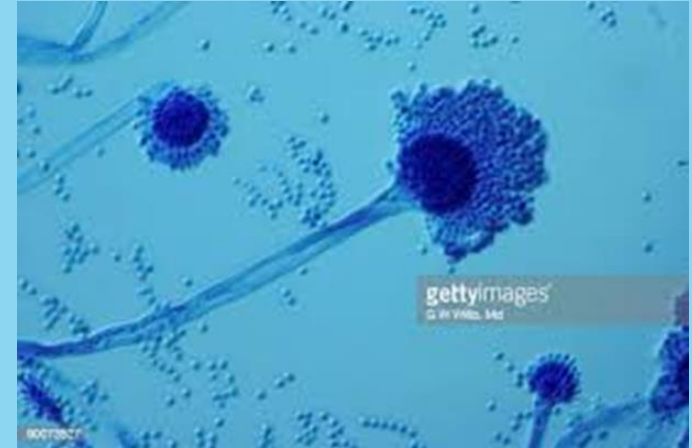
~~Aspergillus fumigatus~~



Aspergillus lentulus



~~Aspergillus flavus~~



Aspergillus caelatus

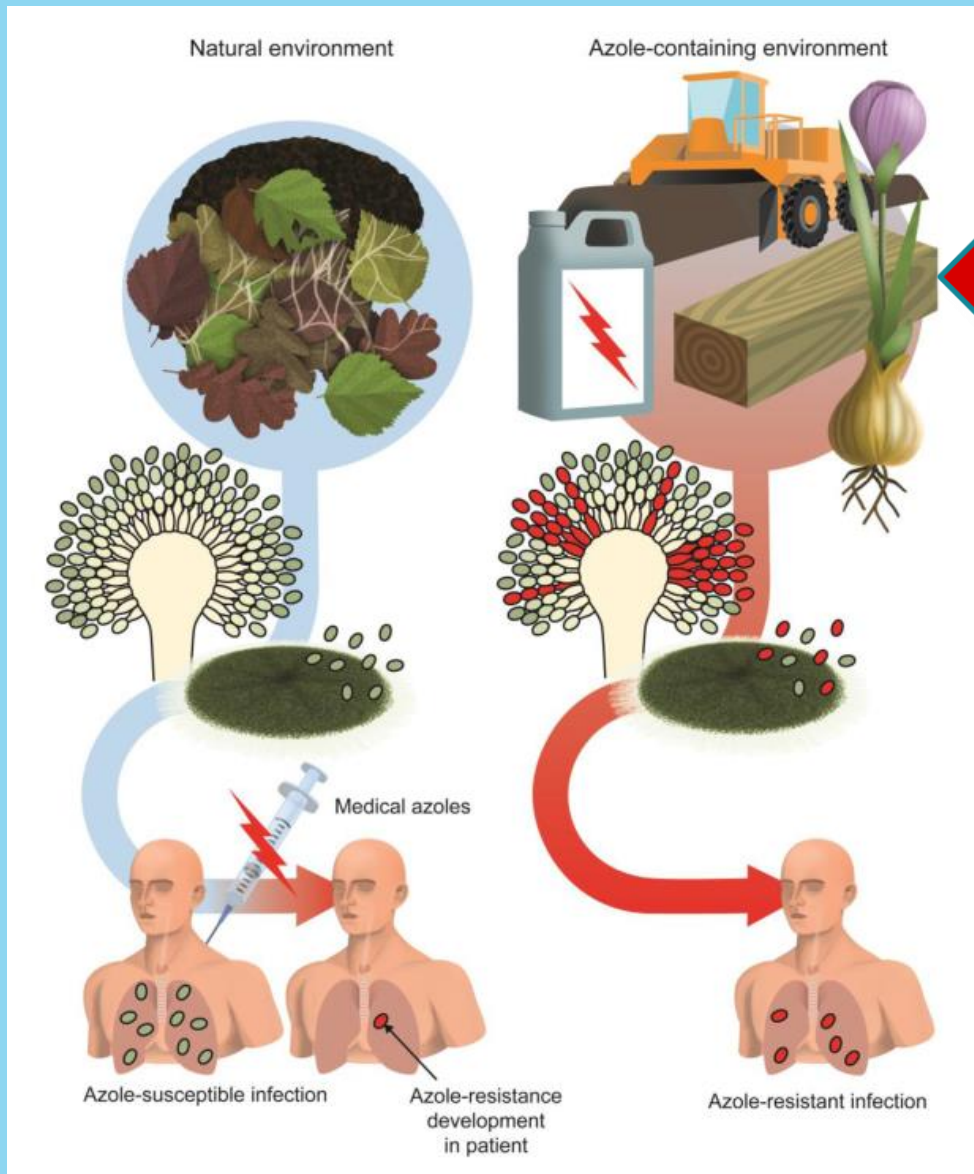
Mode of transmission and Portal of exit

- The portal of exit is closely related to the mode of transmission.
- Aspergillosis is not transmitted from person to person
- the emphasis on focusing control measures on preventing the exit of that organism from the infected patient does not appear to a major component.

Reservoir

▷ **Two main environmental sources:**

- Outside air entering hospital through gaps in filters, windows, backflow of contaminated air
- ▷ Moist environments (e.g., plumbing, leaks, rainwater, air conditioning condensate)



Azole-resistant *Aspergillus*

- * The Netherlands was the European country using the greatest amount of azole fungicide, followed by Germany and France.
- * In fact, the first environmental pan-azole-resistant *Aspergillus fumigatus* isolate was detected in the Netherlands [Lass-Flörl, C.; et al. A nationwide passive surveillance on fungal infections shows a low burden of azole resistance in molds and yeasts in Tyrol, Austria. *Infection* **2018**, *46*, 701–704.].
- * For clinical isolates, a study performed in a cardiothoracic center in London detected a higher prevalence of azole-resistant *Aspergillus fumigatus* (13.2%) principally associated with the environmentally driven TR34/L98H mutation.

Epidemiology of Azole-resistant *Aspergillus*

- ★ Many studies have principally focused on azole-resistant *A. fumigatus* isolates because they represent the predominant pathogen of aspergillosis.
- ★ The first case of ARAF was reported in the late 1980s in the Netherlands.
- ★ The overall azole resistance rate of *A. fumigatus* was reported as ranging from 0.6 to 27.8%, depending on the isolation country, the type of disease, and the emergence of the environmental resistance mechanism.
- ★ Most of the environmental azole-resistant isolates were found in Europe (56.7%) than in other countries due to the higher azole fungicide application per hectare of agricultural land (Burks, C.; Darby, A.; Gomez Londoño Momany, M.; Brewer, M.T. Azole-resistant *Aspergillus fumigatus* in the environment: Identifying key reservoirs and hotspots of antifungal resistance. *PLoS Pathog.* **2021**, *17*, e1009711.).

Prevalence of azole-resistant *Aspergillus fumigatus* strains in European countries.

Pathogens **2023**, 12, 1305.



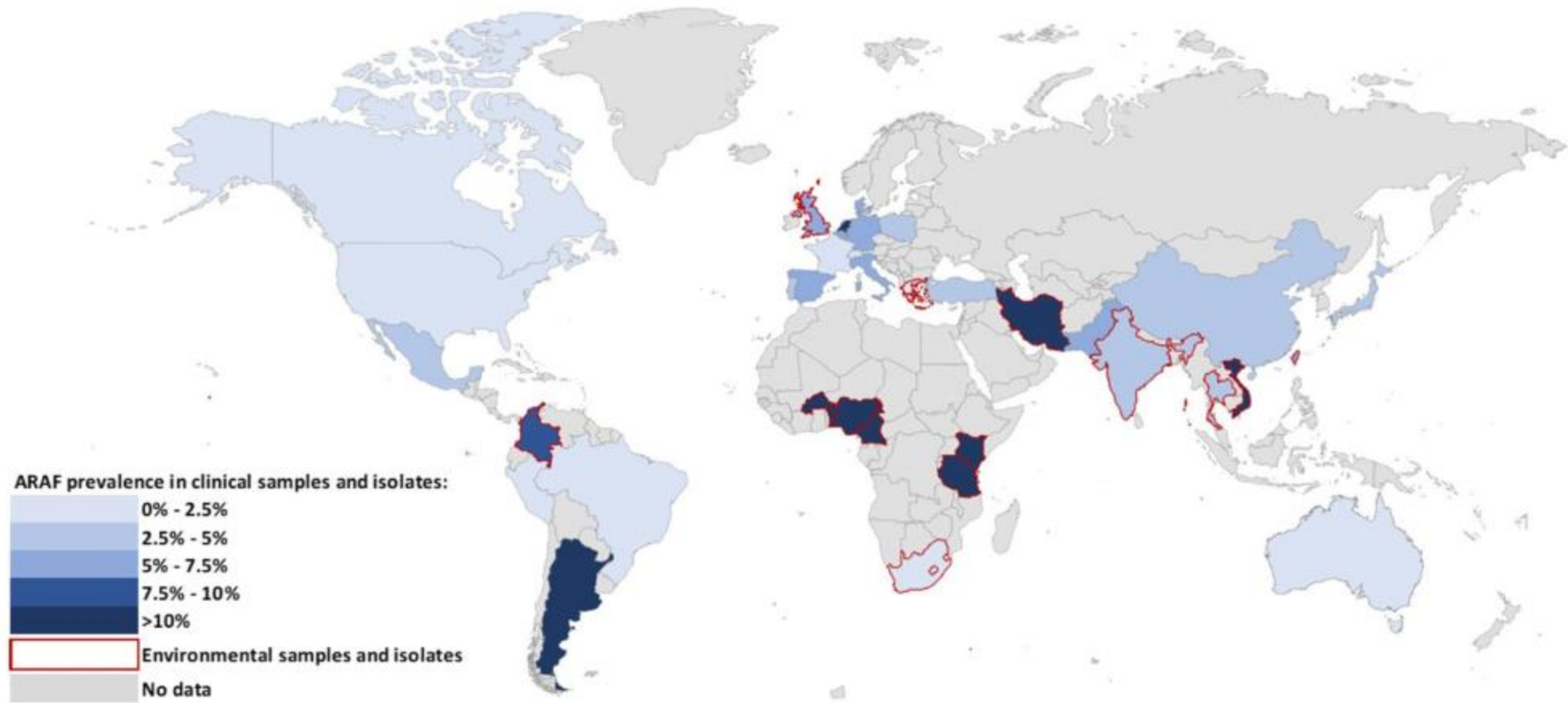


Fig. 1 Worldwide prevalence of azole resistant *Aspergillus fumigatus* (ARAF) clinical and environmental samples and isolates

A note!

- ▷ In a recent study [Fakhim, H.; et al. Trends in the prevalence of amphotericin B resistance (AmBR) among clinical isolates of *Aspergillus* species. J. Mycol. Med. 2022, 32, 101310], among 26,909 *Aspergillus* isolates analyzed, resistance to amphotericin B was detected in 36.8% of *A. terreus*, 14.9% of *A. flavus*, 5.2% of *A. niger*, and 2.01% of *A. fumigatus* isolates.
- ▷ Some *A. lentulus* and *A. ustus* isolates have been reported to show amphotericin B resistance.
- ▷ Additionally, an increasing trend in amphotericin B resistance was observed in *A. fumigatus* between 2016 and 2020, together with a decreasing trend in amphotericin B resistance in *A. terreus* and *A. flavus*.

Range 6.9%- 55.2%

Range 0-11.9%

14.3% *A. fumigatus* isolate resistant to voriconazole

2023

4.1% and 6.1% of *A. fumigatus* isolates resistant to VOR and NWT for ITR. 2% and 11.9% *A. flavus* isolates resistant to VOR and NWT for ITR.

55.2% and 6.9% *A. fumigatus* isolates were NWT and resistant for ITR and VOR respectively. 6.9% had cross resistant to ITR and VOR.

2022

No Azole resistance in 55 *A. fumigatus* clinical isolates were found

12.9% of *A. fumigatus* isolates were NWT and resistant for ITR and VOR. 12.9% isolates had cross resistant to ITR and VOR.

2018

8% of *A. fumigatus* isolates NWT for itraconazole.

8.9% *A. fumigatus* isolates were NWT for ITR and 5.1% isolates were resistant to VOR.

2016

3.5% *A. fumigatus* isolates cross resistant to ITR and VOR.

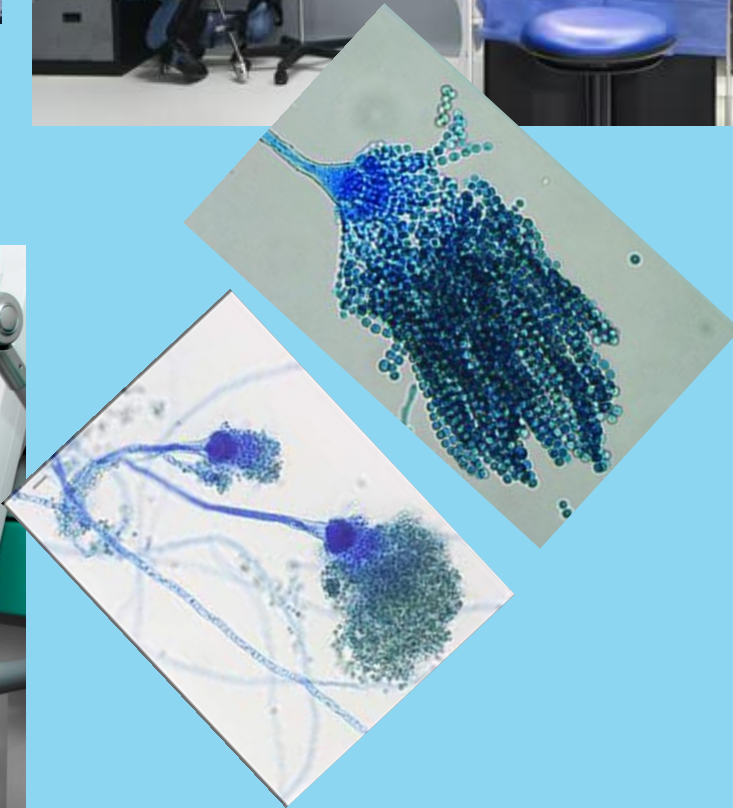
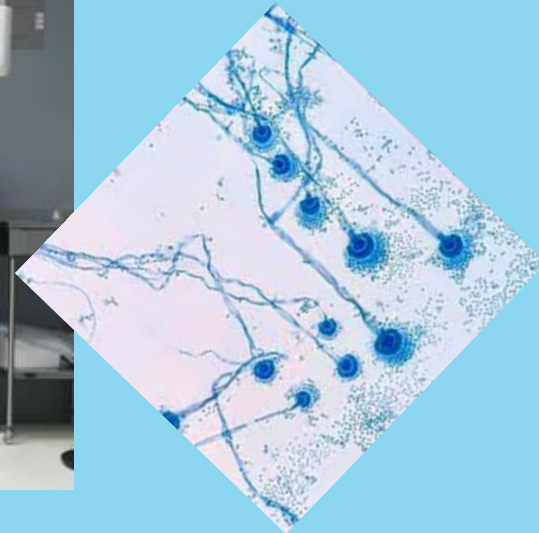
12.2% *A. fumigatus* isolates cross resistant to ITR and VOR

2013

3.2% *A. fumigatus* isolates cross resistant to ITR and VOR.

Environment isolates

Clinical isolates



Available online at www.sciencedirect.com

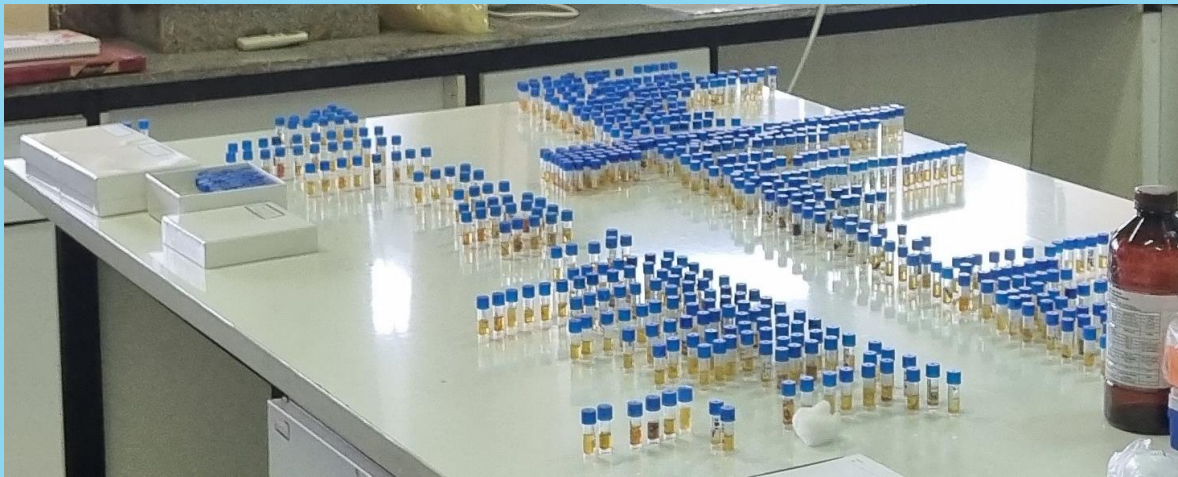
Journal of Hospital Infection

journal homepage: www.elsevier.com/locate/jhin



Electronic equipment and appliances in special wards of hospitals as a source of azole-resistant *Aspergillus fumigatus*: a multi-centre study from Iran

M. Ghazanfari^{a,b}, M. Abastabar^{a,b}, I. Haghani^a, F. Kermani^b, N. Keikha^c,
M. Kholoujini^d, M.H. Minooeianhaghghi^e, S.A. Jeddi^f, A. Shokri^g, A. Ghojoghi^h,
K. Amirizadⁱ, M. Azish^j, Y. Nasirzadeh^b, B. Roohi^b, M. Nosratabadi^{b,k},
S. Hedayati^l, S. Ghanbari^{b,l}, R. Valadan^m, M.T. Hedayati^{a,b,*}



Results

- A total of 693 samples were collected from electronic medical equipment and appliances in 23 educational hospitals from 18 provinces of Iran.

Table I

Distribution of isolated *Aspergillus* species (N=89) on azole containing agar plates

Subgenus	Section	Species	No. of isolates
<i>Fumigati</i>	<i>Fumigati</i>	<i>A. fumigatus</i>	37 (41.6%)
<i>Circumdati</i>	<i>Flavi</i>	<i>A. flavus</i>	5 (5.6%)
		<i>A. oryzae</i>	4 (4.5%)
<i>Circumdati</i>	<i>Nigri</i>	<i>A. tubingensis</i>	21 (23.6%)
		<i>A. niger</i>	14 (15.7%)
		<i>A. welwitschiae</i>	2 (2.2%)
		<i>A. luchuensis</i>	2 (2.2%)
		<i>A. japonicus</i>	2 (2.2%)
		<i>A. awamori</i>	1 (1.1%)
		<i>A. terreus</i>	1 (1.1%)

In Vitro Antifungal Susceptibility Test

- Among different species of *Aspergillus*, *A. fumigatus* showed the highest MIC₅₀/MIC₉₀ to all azoles tested (VCZ: 2/>16 µg/mL; ICZ: 1/16 µg/mL; PCZ: 0.25/1 µg/mL).
- VCZ showed the highest *in vitro* antifungal activity against *A. flavus*
- Whereas *A. niger* had the highest *in vitro* susceptibility to PCZ and ICZ.
- MICs 4, 2, and 1 (µg/mL) were observed as the highest MICs of *A. fumigatus* to VCZ, ICZ, and PCZ, respectively.
- A proportion of 12.8% (89/693) azole resistance *Aspergillus* species were recovered.
- VCZ resistance *A. fumigatus* were recovered as 5.3% (37/693).

Cont. *In Vitro* Antifungal Susceptibility Test

- In terms of the defined MIC breakpoint and ECV for azoles against *Aspergillus* spp. isolates, among the 37 *A. fumigatus* isolates obtained from ACAP, 8.1% were classified as intermediate, and 51.3% were classified as resistant to VCZ.
- The non-wild type of *A. fumigatus* against ICZ was identified in 56.8% of isolates.
- The non-wild type of *A. niger* to VCZ was observed in 7.1% of isolates.
- One isolate of *A. flavus* (1/5, 20%) was reported as non-wild type to PCZ.
- Of the 37 isolates of *A. fumigatus*, 48.6% showed cross-resistance to VCZ and ITZ.

Table III
 Characteristics of azole-resistant and non-wild-type *A. fumigatus* isolates and detected mutations

Isolate number	Isolated from ACAP			MIC ($\mu\text{g}/\text{mL}$)			<i>CYP51A</i> mutations											
	VCZ	ICZ	PCZ	VCZ	ICZ	PCZ	F46Y	G54E	G138C	Y121F	M172V	M220I	D255E	T289F	G432C	G448S	L98H	TR34
22s		+		4	8	0.063	+	+	+	-	+	+	+	+	+	+	-	-
3	+			4	4	1	+	+	+	-	+	+	+	+	+	+	+	+
4	+			2	4	0.063	+	+	+	-	+	+	+	+	+	+	+	+
14	+			>16	2	0.5	+	+	+	+	+	+	+	-	+	+	-	-
15	+			4	16	1	+	+	+	-	+	+	+	+	+	+	+	+
18	+			4	16	0.5	+	+	+	-	+	+	+	+	+	+	+	+
19	+			4	>16	1	+	+	+	-	+	+	+	+	+	+	+	+
21	+	+		4	16	1	+	+	+	-	+	+	+	+	+	+	+	+
29	+			>16	2	0.125	+	+	+	+	+	+	+	-	+	+	-	-
35	+			4	8	2	+	+	+	-	+	+	+	+	+	+	+	+
46	+	+		8	>16	1	+	+	+	-	+	+	+	-	+	+	+	+
47	+	+		4	>16	0.5	+	+	+	-	+	+	+	+	+	+	+	+
56		+		16	2	0.25	+	+	+	+	+	+	+	-	+	+	-	-
63	+	+		>16	2	1	+	+	+	+	+	+	+	-	+	+	-	-
72		+		8	>16	1	+	+	+	+	+	+	+	+	+	+	+	+
97	+			>16	1	0.063	+	+	+	+	+	+	+	-	+	+	-	-
23	+			4	16	1	-	-	-	-	+	+	+	+	+	+	-	-
34	+			2	2	1	+	+	+	-	-	-	-	-	+	+	-	-
65	+			>16	2	0.5	+	+	+	+	+	+	+	-	+	+	-	-

ACAP, azole-containing agar plate; MIC, minimum inhibitory concentration; ICZ, itraconazole; VCZ, voriconazole; PCZ, posaconazole.

High prevalence of Terbinafine resistance among *T. mentagrophytes*/*T. interdigitale* complex isolated from Iran

Iman Haghani^{1,2}, Maryam Babaie¹, Akbar Hoseinnejad^{3,4}, Ali Rezaei-Matehkolaei⁴, Ramin Mofarah⁵, Zahra Yahyazadeh¹, Firoozeh Kermani⁶, Javad Javidnia^{1,2}, Tahereh Shokohi^{1,2}, Maryam Azish⁷, Kambiz Kamyab Hesari⁸, Majid Saeedi⁹, Zeynab Ghasemi⁸, Shaghayegh Khojasteh¹⁰, Zohreh Hajheydari¹¹, Elham Mosayebi^{1,2}, Reza Valadan¹², Seyedmojtaba Seyedmousavi¹³, Mahdi Abastabar^{1,2*}, M.T. Hedayati^{1,2*}



Recommendation

- Active surveillance for potential sources of fungal contamination
- *In vitro* antifungal susceptibility testing for all clinical fungal isolates routinely
- *A. flavus* should be considered as the main cause of IA in Iran
- Testing for identification of mutations in clinical fungal isolates routinely
- Presence of a real infection control committee in all hospitals



Thanks!

hedayatimt@gmail.com

19 February
2024



Dr. M. Ghazanfari

S. Ghanbari

B. Nikoueian

S. Yaalimadad

